

Research note

Incidence of Citrus exocortis viroid and Hop stund viroid in commercial citrus groves from Morocco

Imane BIBI^{1*}, Ez-zahra KHARMACH¹, Zouheir CHAFIK², Jamal BEN YAZID¹, Anwar BOUSAMID², Mohamed AFECHTAL³

¹Laboratory of Biochemistry and Biotechnologies, Faculty of Sciences, Mohammed First University of Oujda, Morocco

²Laboratory of Plant Biology and micro-organisms, Faculty of Sciences, Mohammed First University of Oujda, Morocco

³National Institute for Agricultural Research (INRA), Regional Center of Kénitra, Kénitra, Morocco

*Corresponding author
Bibi.phytia@gmail.com

Received 12/03/2020
Accepted 18/04/2020

Abstract

Viroids are the smallest known pathogens of plants. They are single-stranded, circular, rod-like RNAs with no protein capsid nor any detectable messenger activity. Citrus species are natural hosts of at least seven viroids of the Pospiviroidae family. Exocortis and cachexia are the most important viroid diseases of citrus in Morocco. In order to assess the incidence and distribution of Citrus exocortis viroid (CEVd) and Hop stunt viroid (HSVd), extensive surveys were conducted, between 2008 and 2018, in the main citrus growing areas of the country. A total of 100 commercial citrus groves of different varieties were inspected for symptoms of exocortis and cachexia, and 5390 samples were collected for laboratory analyses, using imprint hybridization. Representative positive and negative samples for each viroid were selected and submitted for additional confirmatory RT-PCR assays using specific primers. The incidence of infection by viroids was about 29.0 %, regardless of citrus species and location, although a slightly higher number of infected trees were located in the Gharb region (41.5 %). CEVd was detected in 11.3 -18.2% of the tested samples, while HSVd was detected up to 10.8 % of the samples tested. Since CEVd and HSVd are readily mechanically transmissible via working tools, they affect both old and young plantings in all the surveyed regions.

Keywords: Citrus exocortis viroid, Hop stunt viroid, citrus, survey, Morocco.

INTRODUCTION

Citrus trees are a major fruit crop in Morocco. They cover about 125,000 ha with an average yearly production of 2 million tons of which about 40 % is exported as fresh fruits (Anonymous, 2017).

Viroids are small infectious agents of single-stranded, unencapsidated, nontranslated, circular RNA with self-complementary sequences (Flores *et al.*, 1997) and cause diseases in economically important crops. These viroids have small genomes ranging from 284 to 375 nucleotides (Duran-Vila *et al.*, 1988). Citrus viroids have been classified into distinct groups based on their biological and physical properties: Citrus exocortis viroid (CEVd) with numerous variants reported (Gross *et al.*, 1982; Visvader and Symons, 1985) and specific citrus variants (referred as CVd-IIb) of hop stunt viroid (HSVd) are causal agents of the exocortis (Semancik and Weathers, 1972) and cachexia (Semancik *et al.*, 1988) diseases, respectively. Three additional viroids (CVd-I, CVd-III and CVd-IV) and another HSVd variant (referred to as CVd-IIa) are known to induce specific symptoms on Etrog citron (*Citrus medica* L.) (Duran-Vila *et al.*, 1988) and CVd-I and CVd-III affect trees grown on trifoliolate orange (*Poncirus trifoliata* (L.) Raf.) rootstock (Roistacher *et al.*, 1993; Semancik *et al.*, 1997). Viroids are graft- or mechanically transmissible agents, disseminated through budding.

Citrus exocortis viroid (CEVd) was described in 1948 as a bark shelling or scaling disorder affecting trees

grown on the trifoliolate orange (*Poncirus trifoliata* (L.) Raf.) rootstock (Fawcett and Klotz, 1948) and is widely prevalent in citrus growing areas of the world (Timmer *et al.*, 2000). CEVd is the largest citrus viroid at 371-nt, and produces the most pronounced symptoms of leaf epinasty, stunting, and necrosis of the leaf midvein. CEVd is a species of the genus *Pospiviroid* and it was the first citrus viroid described (Roistacher, 1991). CEVd is distributed primarily by the introduction and propagation of infected budwood, by top working and by mechanical transmission (Garnsey and Jones, 1967). CEVd is not known to be vector or seed transmitted.

Cachexia disease is found in most citrus growing countries of the world (Roistacher, 1983). Hop stunt viroid (HSVd), a species in the *Hostuviroid* genus, contains 295-303 nucleotides (Roistacher and Garnsey, 1988; Reanwarakorn and Semancik, 1999) and is a major pathogen of mandarin and tangerine species in the world (Catara *et al.*, 1988) causing tremendous losses in yield and quality of the fruits. Cachexia was first described in 1950 as a disease of Orlando tangelo with the following symptoms: discoloration, gumming and browning of phloem tissue, wood pitting and bark cracking (Childs, 1950). The viroid is transmitted by vegetative reproduction or mechanical means, e.g. pruning or other practices, which may lead to plant damage. Mechanical harvesting and other operations lead to spread along the rows in commercial hop plantations (Pethybridge *et al.*, 2008).

The object of this research was to investigate two different species of citrus viroids in Morocco (CEVd and HSVd), via analyzing the samples that showed different symptoms in citrus trees. The work will collect some basic information about incidence and geographical distribution of these two species of citrus viroids in Morocco.

MATERIAL AND METHODS

Field surveys were conducted between 2008 and 2018, in the main citrus growing areas of the country: Gharb, Haouz, Loukkos, Moulouya, Souss and Tadla. A total of 100 commercial citrus groves of different varieties were inspected for symptoms of exocortis and cachexia, and 5390 samples were randomly collected from different mature trees (15- to 20-yr-old) of sweet orange (*Citrus sinensis*) cvs, Washington Navel, Valencia late, clementine (*C. clementina*) and lemon (*Citrus limon*). Plants showing symptoms of viroids were also included in the sampling. All samples were tested by imprint hybridization for Citrus exocortis *pospiviroid* (CEVd) and Hop stunt *hostuviroid* (HSVd) according to the protocol described by Palacio *et al.*, (1999, 2000). For confirmatory tests, representative positive and negative samples for each viroid were selected and submitted for additional RT-PCR assays using specific primers for each agent (Wang *et al.*, 2009). Total nucleic acids (TNAs) were recovered from 0.2 g of leaf petioles ground in liquid nitrogen as described by Foissac *et al.*, (2001). For reverse transcription, first strand cDNA was synthesized from 0,5 µg of total RNA using random primers (Noris *et al.*, 1994).

RESULTS AND DISCUSSION

The results of the present survey showed that HSVd and CEVd were found to be widespread with an infection rate of 10.8 % and 18.2 % of the tested samples and were detected in almost all citrus species and cultivars (Table 1). CEVd and HSVd were detected in most surveyed groves across the country, but not in the nurseries. Typical symptoms of bark cracking and scaling of exocortis were observed on Citrange Carrizo rootstock in the Gharb, Loukkos and Tadla regions. The highest infection rate of CEVd was recorded in the Gharb region with an infection rate of 29% followed by Loukkos region with 20%. Cachexia was frequently found in mandarin orchards in most of the country's citrus production areas. In the nurseries surveyed, No HSVd-infected plant was detected. Afechtal *et al.*, (2016) found the Hop stunt viroid on common clementine and Valencia late in the Gharb

region. Most of these inspected trees carried more than one viroid. In most cases, the infected trees are asymptomatic carriers because sour orange, the predominant rootstock used in Morocco, does not show symptoms of viroid infection.

Similar results were reported by Malfitano *et al.*, (2005) in Campania (Italy) who found that HSVd and CEVd are the most widespread viroids infecting 87 % and 68 % of the tested samples and were detected in almost all citrus species and cultivars. They were frequently found in mixed infection. In Tunisia, HSVd and CEVd were described by Duran-Vila *et al.*, (1988) by molecular hybridization. Their results showed that these two viroid species were highly widespread accounting respectively for 70.3% and 72.3 % of the sources tested.

In the present study, imprint hybridization has been applied for the evaluation of occurrence and distribution of citrus viroids in Morocco. A total of 20 representative positive and negative samples for viroids (CEVd, and HSVd) were selected and submitted for additional RT-PCR assays using specific primers for each viroid. In the assay, positive and negative controls were included, for each agent. RT-PCR results were similar to those obtained above using molecular hybridization technique. Both techniques confirmed the presence of the CEVd and HSVd in all surveyed areas.

The high incidence of CEVd and HSVd observed during the survey is probably due to the use of infected bud wood material. Therefore, the use of healthy (virus-free) planting material, usually within the framework of a phytosanitation program based on diagnosis, detection and elimination of the causal agent(s) through sanitation could prevent viroid infection. Contamination tools and top-grafting may have been responsible for viroid spread and accumulation in individual trees. Therefore, transmission and dissemination of viroids could be prevented by treating budding knives, pruners, clippers in sodium hypochlorite solution (Arif *et al.*, 2005). Furthermore, it is important to mention that the high viroid contamination rate should be taken into consideration when new rootstocks will be chosen to manage the tristeza disease.

The citrus samples of different cultivars tested in this study were from major citrus growing regions, therefore, represent a wide geographical distribution and host diversity of citrus viroids. It's the first report on the occurrence and distribution of citrus viroids on citrus plants in Morocco and this investigation will provide preliminary characterizations for further research of citrus viroids in Morocco.

Table 1: Detection of citrus viroids in Morocco

Region	Tested trees No.	CEVd		HSVd	
		No.	%	No.	%
Gharb	1000	290	29.0	125	12.5
Haouz	600	132	22.0	66	11.0
Loukkos	550	110	20.0	42	7.8
Moulouya	760	135	17.8	150	19.7
Souss	1860	211	11.3	119	6.4
Tadla	620	102	16.4	79	12.7
Total	5390	980	18.2	581	10.8

REFERENCES

- Afechtal M., Jamai H., Mokri F., Essarioui A., Fadoul Z., Sbaghi M., Dababat A. A. (2016). First Report of *Hop stunt viroid* Infecting Citrus Trees in Morocco. *Plant Disease*, 100: 1512.
- Arif M., Ahmad A., Ibrahim M., Hassan S. (2005). Occurrence and distribution of virus and virus-like diseases of citrus in north-west frontier province of Pakistan. *Pakistan Journal of Botany*, 37: 407.
- Catara A., Azzaro A., Mughal S. and Khan D. (1988). Virus, viroid and prokaryotic diseases of citrus in Pakistan. In: (eds). *Proceedings of 6th Conference of the International Organization of Citrus Virologist. Tel Aviv*.
- Childs J. (1950). The cachexia disease of Orlando Tangelo. *Plant Disease Reporter*, 34(10).
- Duran-Vila N., Roistacher C., Rivera-Bustamante R. and Semancik J. (1988). A definition of citrus viroid groups and their relationship to the exocortis disease. *Journal of General Virology*, 69: 3069-3080.
- Duran-Vila, N., Roistacher, C. N., Rivera-Bustamante, R., Semancik, J. S. (1988). A definition of citrus viroid groups and their relationship to the exocortis disease. *Journal of General Virology*, 69: 3069-3080.
- Fawcett H. S. and Klotz L. (1948). Exocortis of trifoliolate orange. *Citrus leaves*, 28: 9.
- Flores R., Di Serio F. and Hernández C. (1997). Viroids: the noncoding genomes. In: (eds). *Seminars in Virology*. Elsevier. 8.
- Foissac X., Svanella-Dumas M.J., Candresse T., Gentil P. (2001). Polyvalent detection of fruit tree tricho, capillo and foveaviruses by nested-PCR using degenerated and inosine containing primers. *Acta Horticulturae*, 550: 37-44.
- Garnsey S., Jones J. (1967). Mechanical transmission of exocortis virus with contaminated budding tools. *Plant Dis. Repr*, 51: 0-4.
- Gross H. J., Krupp G., Domdey H., Raba M., Jank P., Losow C., Alberty H., Sanger H. L. and Ramm K. (1982). Nucleotide sequence and secondary structure of citrus exocortis and *chrysanthemum* stunt viroid. *European Journal of Biochemistry*, 121: 249-257.
- Malfitano M., Barone M., Duran-Vila N., Alioto D. (2005). Indexing of viroids in citrus orchards of Campania, Southern Italy. *Journal of Plant Pathology*, 87: 115-121.
- Maniatis T., Fritsh E.F., Sambrook J. (1989). *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory, New York.
- Norris D. P., Patel D., Kay G. F., Penny G. D., Brockdorff N., Sheardown S. A., Rastan S. (1994). Evidence that random and imprinted Xist expression is controlled by preemptive methylation. *Cell*, 77: 41-51.
- Palacio A., Foissac X., Duran-Vila N. (2000). Indexing of citrus viroids by imprint hybridization: comparison with other detection methods. *Proceedings of the Fourteenth International Organization of Citrus Virologists IOCV*, Riverside, CA: 294-301.
- Palacio A., Foissac X., Duran-Vila N. (1999). Indexing of citrus viroids by imprint hybridization. *European Journal of Plant Pathology*, 105: 897-903.
- Pethybridge S. J., Hay F. S., Barbara D. J., Eastwell K. C., Wilson C. R. (2008). Viruses and viroids infecting hop: Significance, epidemiology, and management. *Plant Disease*, 92: 324-338.
- Reanwarakorn K., Semancik J. (1999). Correlation of hop stunt viroid variants to cachexia and xyloporosis diseases of citrus. *Phytopathology*, 89: 568-574.
- Roistacher C. N. (1991). *Graft-transmissible diseases of citrus: Handbook for detection and diagnosis*. FAO.
- Roistacher C.N., S.M. Garnsey. (1988). Cachexia. In: *Compendium of Citrus Diseases..* (Eds.): J.O. Whiteside, S.M. Garnsey & L.W. Timmer. *Amer. Phytopathol. Soc. St. Paul, Minnesota, USA*. 38-39.
- Roistacher C., Bash J., Semancik J. (1993). Distinct disease symptoms in *Poncirus trifoliata* induced by three citrus viroids from three specific groups. In: (eds). *International Organization of Citrus Virologists Conference Proceedings (1957-2010)*. 12.
- Roistacher C., Gumpf D., Nauer E., Gonzales R. (1983). Cachexia disease: virus or viroid. *Citrograph*, 68: 111-113.
- Romero-Durbán J., Cambra M., Duran-Vila N. (1995). A simple imprint-hybridization method for detection of viroids. *J. Virol. Methods*, 55: 37-47.
- Semancik J., Weathers L. (1972). Exocortis disease: Evidence for a new species of "infectious" low molecular weight RNA in plants. *Nature New Biology*, 237: 242-244.
- Semancik J. S., Morris T. J., Weathers L. G., Rordorf G. F., Kearns D. R. (1975). Physical properties of a minimal infectious RNA (viroid) associated with the exocortis disease. *Virology*, 63: 160-167.
- Semancik J., Rakowski A., Bash J., Gumpf D. (1997). Application of selected viroids for dwarfing and enhancement of production of 'Valencia' orange. *Journal of Horticultural Science*, 72: 563-570.
- Semancik J., Roistacher C., Rivera-Bustamante R., Duran-Vila N. (1988). Citrus cachexia viroid, a new viroid of citrus: relationship to viroids of the exocortis disease complex. *Journal of General Virology*, 69: 3059-3068.
- Timmer L.W., Garnsey S.M., Graham J.H. (Eds.). (2000). *Compendium of citrus diseases. (Second Ed.)*. *Amer. Phytopathol. Soc., USA*.
- Visvader J.E., Symons R. H. (1985). Eleven new sequence variants of citrus exocortis viroid and the correlation of sequence with pathogenicity. *Nucleic acids research*, 13: 2907-2920.
- Wang X., Zhou C., Tang K., Zhou Y., Li Z. (2009). A rapid one-step multiplex RT-PCR assay for the simultaneous detection of five citrus viroids in China. *European Journal of Plant Pathology*, 124: 175-180
- Yang X., Hadidi A., Garnsey S.M. (1992). Enzymatic cDNA amplification of citrus exocortis and cachexia viroids from infected citrus hosts. *Phytopathology* 82: 279-285.