

Marker assisted selection in plant breeding

Fatima HENKRAR

Sripada UDUPA

International Center for Agricultural Research in the Dry Areas (ICARDA), Rabat, Morocco International Center for Agricultural Research in the Dry Areas (ICARDA), Rabat, Morocco

Marker assisted selection (MAS) is 'smart breeding' or fast track plant breeding technology. It is one tool utilized in breeding companies and research institutes for fast development of improved varieties, giving possibility to select desirable traits more directly using DNA markers. In this review, we discussed the use of MAS in biotic, abiotic, quality and other agronomic traits. Besides, we emphasized the importance of MAS at ICARDA and underlined the successful application of MAS in the last 10 years. The use of molecular markers makes the process of selecting parental lines more efficient based on genetic diversity analysis. It can aid the conventional breeding, especially for certain biotic and abiotic traits laborious to manage. Still, MAS contributed very little to the release of improved cultivars with greater tolerance to abiotic stresses, with only a few exceptions. MAS was extensively used to improve rice varieties, mainly resistant to bacterial blight and blast disease and was applied in drought tolerance along with GPC (Grain protein content) in quality traits. MAS at ICARDA is used to characterize new parental materials for disease resistance genes as well as in screening advanced lines with a focus on association mapping and identification of new QTLs. The application of MAS increased in the last decade. It is more and more used in different crops. However, rice is still the dominant crop in terms of number of publications using MAS.

Keywords: marker assisted selection, plant, biotic stress, abiotic stress, quality, ICARDA

Introduction

Wheat breeders continuously seek for new techniques which can be used for assembling target traits into new wheat cultivars and achieve the same breeding progress in a much shorter time than through conventional breeding. The main goals of wheat breeding are increasing the yield, improving the resistance to abiotic and biotic stresses, improving the quality. While simple traits can easily be detected, other complex traits such as disease resistance or drought tolerance are much more difficult to determine for the breeder. Young (1999) wrote: "Before the advent of DNA marker technology, the idea of rapidly uncovering the loci controlling complex, multigenic traits seemed like a dream''. Now with DNA marker technology, this dream became reality. The capacity of DNA markers to detect allelic variation in the genes underlying traits offers a great promise for plant breeding. By using DNA markers to assist in plant breeding, efficiency and precision could be greatly increased. The use of DNA markers in plant breeding is called marker-assisted selection (MAS).

Definition of MAS

Marker assisted selection (MAS) is 'smart breeding' or fast track plant breeding technology. It is one tool utilized in breeding companies and research Institutes for fast development of improved varieties, giving possibility to select desirable traits more directly using DNA markers. The molecular markers can then be used to assist breeders track whether the specific gene or chromosome segment(s) known to affect the phenotype of interest is present in the individuals or populations of interest. The potential of MAS, thus, moving from phenotype based towards genotype based selection using markers linked to gene of interest. Thanks to the advent of DNA



markers in the late of 1970s, it has now become possible to directly target genomic regions that are involved in the expression of traits of interest.

The history of Marker assisted selection

The idea of MAS begins with the theory of quantitative trait loci (QTLs) mapping described by sax (1923), when he observed an association between monogenic trait (Seed coat pigmentation) and polygenic trait (seed size). This concept was further elaborated by Thoday (1961), who suggested mapping and characterizing all QTLs involved in complex traits using single gene marker. The first DNA-based genetic markers were restriction fragment length polymorphisms, RFLPs (Botstein et al., 1980). Permit to construct the first map for tomato using 57 RFLPs in 1986 (Bernatzky and Tanksley, 1986). Beckmann and Soller (1986) described the first use of restriction fragment length polymorphism (RFLP) markers in crop improvement including theoretical issues related to markerassisted backcrossing (MABC) for improvement of gualitative traits. Tanksley et al. (1989) published the use of RLFP as tool to select desirable lines. He reported the possibility to analyzing plants at the seedling stage, screening multiple characters that would normally be epistatic with one another, minimizing linkage drag, and rapidly recovering a recurrent parent's genotype. At that time, the idea of selection of target genes based on genotypes rather than phenotype was extremely attractive to plants breeders (Young, 1999). All those initiatives open the door to marker technology and development of simpler DNA marker involving PCR techniques such as Random-Amplified Polymorphic DNAs, RAPDs (Williams et al., 1990), Amplified Fragment Length Polymorphisms, AFLPs (Vos et al., 1995), Simple Sequence Repeat, SSR also known microsatellites (Powell et al., 1996) and Single Nucleotide Polymorphisms, SNPs (Gupta et al., 2001). Along with, the research boost in DNA marker technology and produce specifics markers like Sequence Characterized Amplified Region, SCAR (Paran and Michelmore, 1993), Cleaved Amplified Polymorphic Sequence, CAPS (Maeda et al., 1990), Sequence Tagged Site, STS (Olsen et al., 1989), Expressed Sequence Tags, EST (Jongeneel, 2000), and most recent marker Diversity Arrays Technology, DArT (Jaccoud et al., 2001).

The application of molecular marker in parental selection and predicting heterosis

The plant breeders seek ways of facilitating the use of available germoplasm effectively for plant improvement. One hand, the use of molecular markers makes the process of selecting parental lines more efficient. Based on genetic diversity calculated from fingerprinting data, plant material can be classified into genetic pools. This information can be extremely helpful for identifying the most appropriate parental lines to be crossed. Lombardi et al. (2014) reported that a selection of divergent parental genotypes for breeding should be made active on the basis of systematic assessment of genetic distance between genotypes, rather than passively based on geographical distance. In other hand, classify parental lines into heterotic groups for the creation of predictable hybrids (Acquaah, 2012). The concept of heterotic groups was developed by Maize research using RFLP-based genetic distances of inbreds for the prediction of hybrid performance and heterosis of single crosses in maize has given different results (Melchinger, 1993). The genetic distance estimates based on molecular marker estimates have been effective in grouping related germplasm (Melchinger et al., 1998). Martin et al. (1995) used both pedigree records and Sequence Tagged Sites (STS) molecular markers to determine the relationship between genetic diversity and agronomic performance of the hybrids and they found significant associations between genetic distance based on pedigree and heterosis were found for kernel weight and protein concentration. Zhao et al. (2008) and others also suggested that genetic distances revealed by molecular markers were highly and positively correlated with heterosis in rice. However, the relationship between parents and genotypic variance components in their progenies has been reported as weak or nonsignificant across many studies (Helms et al., 1997; Burkhamer et al., 1998; Melchinger et al., 1998; Bohn et al., 1999; Gumber et al., 1999; Brachi et al., 2010; Hung et al., 2012).



MAS in disease resistance breeding

Plant diseases are the result of infection by other organisms that adversely affect the growth, physiological functioning and productivity of a plant. Plant diseases can drastically affect a country's economy. Therefore, disease management has always been one of the main objectives of any crop improvement program. There are at least 50000 diseases of economic plants and new diseases are discovered every year (Lucas, 1992). Plant diseases are sometimes grouped according to the symptoms they cause (root rots, wilts, leaf spots, blights, rusts, smuts), to the plant organ they affect (root diseases, stem diseases, foliage diseases), or to the types of plants affected (field crop diseases, vegetable diseases, turf diseases, etc.) (Agrios, 2004). Using plant resistance genes for developing disease-resistant varieties are a convenient alternative to other measures like pesticides or other chemical control methods employed to protect crops from diseases (Gururani et al., 2012). That is the objective of plant breeding, the identification of resistant plants, which are then crossed with agronomically acceptable but susceptible plants. A program of backcrossing to the susceptible parent and selection of resistant phenotypes leads to the production of plants that are similar to the susceptible parent but having the required resistance. Breeders have successfully developed lines resistant to disease by integrating R-genes into their cultivars. However, it is not always the case due to the time-consuming by conventional breeding process that take around 10 years, and by this time, in some instances, the pathogen has already evolved a variant that is not recognized by the improved cultivar, leading to susceptibility. DNA markers have enormous potential to improve the efficiency and precision of conventional plant breeding via marker-assisted selection (MAS) by reducing the reliance on laborious and fallible screening procedures. Especially for durable resistance or no specific, that becomes a challenge and the best way to overcome the new races pathogen evolution. The use of molecular markers in selection can aid the conventional breeding, especially for certain traits laborious to manage it. Xu and Crouch (2008) specify four kinds of traits which DNA markers should be helpful. (i) traits that are difficult to manage through conventional phenotypic selection because they are expensive or time-consuming to measure, have low penetrance or complex inheritance; (ii) traits whose selection depends on specific environments or host developmental stages; (iii) maintenance of recessive alleles during backcrossing or for speeding up backcross breeding in general; and (iv) pyramiding multiple monogenic traits or several QTL for a single disease resistance with complex inheritance. Several studies reported the application of molecular markers as a tool to assist phenotypic method to improve concerned traits. For example, Miklas et al., (2006) reported in bean that the most effective strategy to improve bean host plant resistance to common bacterial blight was a combination of MAS with periodic phenotypic selection, because it allows the retention of minor QTL and selects epistatic interactions that contribute to improved disease resistance. Wilde et al. (2008) noted the efficiency of MAS with phenotypic selection combination in improving resistance against Fusarium head blight. One of the successful applications of MAS in breeding disease resistance was in Indonisia, and the release of two rice varieties 'Angke' and 'Conde', which are resistant to bacterial leaf blight infection (Bustamam et al., 2002). Also, Zhao et al. (2012) succeed in introgression of gHSR1, which is a OTL related to head smut in head smut-susceptible lines via marker-assisted selection, which has significantly reduce disease incidence over time in maize.

MAS in abiotic stress breeding

Abiotic stress is defined as environmental conditions that reduce growth and yield below optimum levels. Plant responses to abiotic stresses are dynamic and extremely complex (Cramer, 2010; reviewed by Cramer et al., 2011). Boyer (1982) indicated that environmental factors may limit crop production by as much as 70%. Many genes affect stress tolerance, but few of the identified genes have proven useful in the field. The genomics era has allowed dissection of the physiological and molecular traits underlying stress tolerance mechanisms to an unprecedented level. Integrated omics analyses have markedly increased our understanding of plant responses to various stresses. These analyses are important for comprehensive analyses of abiotic stress responses, especially the final steps of stress signal transduction pathways (Cramer et al., 2011). The application of omics



technologies has contributed to the development of stress-tolerant crops in the field. Several genes are identified to have a great role in abiotic stress tolerance. For instance, SNACs were characterized as factors that regulate expression of genes important for drought and salinity tolerance in rice (Hu et al., 2006; reviewed by Todaka et al., 2012). DREB1/CBF regulon involved in cold-stress-responsive gene expression, and DREB2 involved in osmotic-stress-responsive gene expression (Yamaguchi-Shinozaki and Shinozaki, 2006). The reviews of Nakashima et al. (2009) and Todaka et al. (2012) discussed more about different abiotic stress genes identified in transcriptomic analyses. This comprehensive knowledge about the genes involved in stress response and tolerance will further allow a more precise use of MAS and transgenics (Dita et al., 2006). However, still MAS contributed very little to the release of improved cultivars with greater tolerance to abiotic stresses, with only a few exceptions (LeDeaux et al., 2006; MacMillan et al., 2006; Ribaut and Ragot, 2007; Welcker et al., 2007). The marker assisted selection was applied especially in drought tolerance. For instance, Courtois et al. (2003) used MAS to transfer a number of QTLs related to a deep rooted character from the japonica upland cultivar "Azucena" to the lowland indica variety "IR64". MAS selected lines showed a greater root mass and higher yield in drought stress. Steele et al. (2004) made novel method termed Marker-evaluation selection in rice crop. This approach used a very large segregating population derived from a wide cross between the upland variety Kalinga III and the irrigated variety IR64. The population was selected for overall agronomic performance in several target stress environments over many generations and the products from the selection were evaluated with markers. Varieties developed through MABC (e.g. Ashoka 228) have better drought resistance as they yield more than parent Kalinga III. Similarly, Steele et al. (2006) used marker assisted breeding program to improve some root traits related to drought tolerance in an Indian rice cultivar Kalinga III. They introgressed five QTL regions associated with root traits from Azucena into Kalinga III. The target OTL on chromosome 9 (RM242-RM201) significantly increased root lengths under drought stress.

MAS in improving agronomic and seed quality traits

Development of cultivars with high agronomic performance and good quality is preeminent in crop breeding programs. Several agronomic and quality traits are polygenic trait controlled by many QTL/genes with smaller effects, such as yield and GPC, seed size seed oil content, days to flower and to maturity, fiber length and strength, etc.; or by few QTL/genes with major effects such as kernel color, flower color, stem color, etc. Those traits cannot be found through phenotypic evaluation alone because they are highly sensitive to environmental changes. In addition, it is difficult to produce ideal cultivars with high yield and good quality due to the existing negative correlation between those traits (Barnard et al., 2002; Chung et al., 2003; Yagdi and Sozen, 2009; Sourour et al., 2018; Ma et al., 2012). Therefore, Molecular detection and genetic tracking of quantitative trait loci (QTL) for agronomic and quality traits will affect positively in manipulation of those traits, and will increase the accuracy of selection. Hence, the identification of QTLs related to quality and agronomic traits is important as an entry point for marker assisted selection. Nowadays, the studies are focusing on desiccation of stable QTLs responsible for agronomic and quality traits in major crops using genome wide association mapping (GWAS), linkage mapping and single nucleotide polymorphism (SNPs). Chen et al. (2016) identifies useful QTL qGW4.05 related to Kernel weight and kernel size in Maize. The agronomic and quality traits of Brassica napus has been dissected using Genome wide association mapping and using a 6K single nucleotide polymorphism (SNP) array (Körber et al., 2016). New QTLs associated with protein and oil content were identified (Cao et al., 2017; Karikari et al., 2019).

The MAS was extensively used for improving GPC. The selection and introgression of a high GPC allele of Gpc-B1 has been achieved in several of the released wheat cultivars (DePauw et al., 2005; Humphreys et al., 2010; Randhawa et al., 2013) using molecular markers. A successful example of an integrated approach of combining phenotypic selection with marker assisted backcross breeding in wheat for introgression of Gpc-B1 in Indian wheat cultivar HUW468 (Vishwakarma et al., 2016). MAS was adopted for studying the genome composition of winter cultivars Zhengmai 7698 using



closely linked or functional markers for gluten protein quality, grain hardness and flour color (Li et al., 2018). It was used to improve oil content in sunflower, and the Marker F4-R1 was validated and proved to be the most efficient in detecting high oil content in sunflower (Dimitrijević et al., 2017). Besides, The MAS was frequently used in the most important trait, yield. Liang et al. (2004) developed a new stable improved line '9311xOryza rufipogon' with yield-enhancing genes and high yield potential using SSRs tightly linked markers. Kumar et al. (2018) combined grain yield and genotypic data from different generations (F3 to F8) for five marker-assisted breeding programs for analyzing the effectiveness of synergistic effect of phenotyping and genotyping in early generations. They found genotyping and phenotyping cost savings of 25–68% compared with the traditional marker-assisted selection approach.

Marker assisted selection at ICARDA

Crop improvement at ICARDA aims to conserve agricultural biodiversity in dry areas and to use these resources to improve food crops through breeding. It covers durum and bread wheat, barley, chickpea, lentil, faba bean, grasspea, and forage and pasture crops. ICARDA's approach combines conventional and biotechnology research to identify molecular markers and to use it. Identification and utilization of molecular markers for marker assisted selection would enhance the development of widely adapted and high yielding varieties with resistance/tolerance to abiotic and biotic resistance and acceptable level of end use quality. The benefit of this 'marker-assisted selection' is that it will make the breeding process faster and more precise. As a result, breeders and farmers will see rapid improvements in crop production, enabling them to improve livelihoods and boost food security.

MAS at ICARDA is used to characterize new parental materials for disease resistance genes (stripe rust, leaf rust, stem rust, nematodes); insect resistance (Hessian fly and Russian Wheat Aphid), phonological traits such as photoperiodism (Ppd), vernalization requirement (Vrn); plant height (Rht), grain hardness and other desirable genes (Tadesse et al., 2012 and 2016). Molecular markers are also used for pyramiding different resistance genes and developing multi-line cultivars targeting for durable resistance to the disease. It helps of screening real hybrids F1, F2, BC1F1 populations. The use of molecular markers and MAS started at ICARDA since long, by identifying and mapping gene resistance to lentil, pea and chickpea pathogen (Baum et al., 2000). The use of molecular techniques and biotechnology tools have expanded considerably, the techniques are applied to almost all crops and concentrated on the development of marker-assisted selection and characterization and identification of fungal pathogens and nematodes. ICARDA has focused on the propagation of the molecular techniques and their application in crop improvement by organizing extensive training to young researchers, students, junior level scientists, and also technicians (Ryan et al., 2012). CIMMYT, Biodiversity, International Centre for Agricultural Research in the Dry Areas (ICARDA), and IRRI have partnered with national research organizations from 13 countries in Africa and South Asia to co-generate and share technologies for genetic characterization and marker-assisted improvement of wheat, barley, and rice, focusing on traits and alleles that are important for the crops adaptation to climatic changes (Halewood et al., 2018). Several works done by ICARDA scientists and students on MAS were published. (Halewood et al., 2018) discriminates between resistant and susceptible chickpea genotypes using two codominant markers associated to Ascochyta blight. Molecular marker associated with grain yield under drought conditions such as the CID, are actively and effectively used in the ongoing breeding program (Nachit, 1998; Nachit and Elouafi, 2004). Dura et al. (2012) identified potential targets for MAS of grain yield improvement in durum wheat in ICARDA laboratory. Recently, the markers assisted selection has been successfully used to enhance tolerance against Barley scald (Sayed & Baum, 2018). Nowadays, ICARDA is focusing on Association mapping (AM) using phenotypic and genotypic data of association panels, due to the importance of this approach in identifying molecular markers (QTLs) linked to traits of interest for potential use in marker assisted selection. In Barley, association mapping was undertaken to identify QTL effective against Psh individual races at seedling stage and QTL for quantitative resistance to barley stripe rust at seedling and adult plant



stages (Visioni et al., 2018). In wheat, genome-wide association mapping (GWAM) was employed using DArT markers technology and ICARDA's elite wheat genotypes to identify markers linked to stripe rust resistance genes in wheat identify closely associated markers with YR resistance for possible use in MAS (Tadesse et al., 2014; Jighly et al., 2015) employed genome-wide association mapping (GWAM). In pulse, the association mapping was designed to determine the genetic basis of seed Fe and Zn concentration in lentil by using single-nucleotide polymorphism (SNP) array derived from cultivated lentil sequences (Singh et al., 2017).

Successful application of MAS in last decade

The MAS of smart breeding method is the method of choice for all breeders. It has been implemented in different crop programs. Several publications declare the application of MAS in crop improvement. But still the number of successful application of this method is less compared to the number of QTLs mapped or markers developed. Moreover, most marker associations are not robust enough for successful marker assisted selection (Young et al., 1999). By using Harzing's Publsih or Perish software (Harzing, 2007) and using the query 'Marker assisted selection' in Google scholar and in Scopus between 2010 and 2019, around 571 publications were retrieved in which the title included 'Marker assisted'. At first sight it was often difficult to distinguish from the title whether a publication is actually reporting a MAS application or if only potential MAS applications of the actual research outputs are discussed. Therefore, the publications were selected by reading the abstracts and sometime the material and methods to distinguish the real application of MAS. The results mentioned in table 1 is the number of publications harvested using MAS keyword. Among 571, only 189 publications were the real applications of MAS. Whereas, others publications were reviews of MAS (163 publications), QTL mapping or identification and/or marker development and validation (149 publications), Characterization and genetic diversity (47 publications) or genomic selection (23 publications). The MAS practical publications were dominant in rice with 87 publications (Figure 1), 29 of them are on bacterial blight diseases. The number of publications in other cereals was limited to 39 publications (18 publications in wheat, 18 publications in Maize and 3 publications in Barley). Marker assisted backcrossing (100 publications, Figure 2) has been most widely and successfully used up-to-date. It has been applied to different crops, e.g. rice, wheat, maize, barley, pear millet, soybean, tomato, etc. compared to other methods such as pedigree method (40 publications), pyramiding (45 publications) and MARS (4 publications).



Table 1: Number and type of publications retrieved from Harzing's Publish from 2010 to 2019 (Harzing, 2007)

Type of publications	Number of publica- tions
MAS articles	189
Reviews	163
Characterization or genetic diversity	47
Mapping or marker development	149
Genomic selection	23
Total	571

Figure 1:



Figure 1: Number of MAS publications applied to different crops in the last 10 years

Figure 1:

Number of publications



Figure 2: Number of publications for different types of MAS collected during the last 10 years

Figure 2:

The successful publications using MAS in last 5 years is resumed in table 2. One successful example of marker assisted backcrossing and pyramiding is the introgression of three BB resistance genes (Xa21, xa13 and xa5) from BB-resistant donor variety IRBB-60 into the BB-susceptible Basmati variety CSR-30 (Baliyan et al., 2018). A successful introgression of Introgression of Shoot Fly (Atherigona soccata L. Moench) Resistance QTLs into Elite Post-rainy Season Sorghum Varieties (Gorthy et al., 2017). An example of a successful application of MAS in breeding new cultivars is the development of "Mura Salad" a new fresh pepper cultivar (Capsicum annuum) containing capsinoids, low-pungent capsaicinoid analogs using dCAPs and SCAR markers (Tanaka et al., 2014). In legumes, a successful application of marker assisted backcrossing in chickpea and specific markers for Fusarium wilt-resistance generate the development of new cultivars Super Annigeri 1 and improved JG 74 with enhanced resistance and improved yielding (Mannur et al., 2019). MAS were successfully applied in wheat to improve GPC-B1 (84-60) and also in Barley to transfer a thermostable β -amylase gene (Xu et al., 2018) scald (Rhynchosporium commune L.) resistance gene (Sayed and Baum, 2018).



Table 2: Examples of successful use of marker assisted selection in different crops for the last 5 years

Target trait	Gene (s)/QTl(s)	Type of Marker used	Name of marker used	Crop	Reference
Blast	Pi2	STS	Pi2-4, HC28	Rice	Yang et al., 2019
Bacterial blight and aroma	Xa21, xa13, xa5, fgr	STS	pTA248, RG136, RG556, BAD2	Rice	Baliyan <i>et al.,</i> 2018
Quality protein	Opaque2 (o2)	SSR	umc1066 and phi057	Maize	Hossain <i>et al.,</i> 2018; Pukalen- thy <i>et al.,</i> 2019
Scald	Rrs1	SSR/ SCAR	Ebmac0871- SSR, HVS3-SCAR, Bmag0006- SSR	Barley	Sayed and Baum, 2018
HMW, Grain hardness, Lipoxygenase,Yellow pigment content, Polyphenol oxidase, Powdery mildew, Yel- low rust, Pre-harvest sprouting		SSR/STS, allele specific	UMN19, Bx7, ZSBy8, ZSBy9a, UMN25, Dx5, UMN26, Pinb-D1a, LOX16, LOX18, YP7A, YP7B-1, YP7D-1, PP018, PP019, PP029, Pm2, Pm4b, Pm8, Xgwm582, Xcfa2040, PHS1, PHS-4AL	Wheat	Li et al., 2018
Blast	Pi54, Pi1 and Pita	STS, SSR	Pi54MAS, RM224, YL155/87	Rice	Khan et al., 2018
Bacterial blight	Xa38, Xa21, Xa13 and Xa5	Gene specific markers/ STS	Os04g53050-1, pTA248, xa13- Prom, 10603-T10Dw	Rice	Yugander <i>et al.,</i> 2018
Bacterial blight	Gm1, Gm4, xa13 and Xa21	SSR	RM1328, RM22550, xa13 prom and pTA248	Rice	Krishnakumar and Kumaravadi- vel 2018
Mosaic virus	RSC4, RSC8, and RSC14Q	SSR	BARCSOYSSR_14_1413, 4 BARCSOYSSR_14_1417, BARCSOYSSR_14_1417, BARCSOYSSR_02_0606, BARCSOYSSR_02_0610, BARC- SOYSSR_02_0616, BARCSOYS- SR_02_0618, Satt334, Sct_033, MY750	Soybean	Wang <i>et al.,</i> 2017
Rust and coffee berry	SH3, SH?, Ck-1	SCAR/ SSR	SP-M16-SH3, BA-124-12K-f, Sat244, BA-48-210R, CaRHvII 2, CaRHvII 3, CaRHvII 5, Sat 207, Sat 235	Coffea	Alkimim et al., 2017
Striga	SG1, SG3, and SG5	SSR	61RM2, SSR-1 and C42-2B	Cowpea	OMOIGUI et al., 2017
Rust	Lr19 and Lr24	SCAR/ SSR	Xwmc221 and SCS1302	Wheat	Singh et al., 2017
Rust	Lr24 and Lr28	SCAR/ SSR	SCS719, SCS1302607, SCS421570 and Xwmc313	Wheat	Kumar et al., 2017
Drought, Striga her- monthica	SNPs	KASP markers	233 SNPs with KASP assay	Maize	Abdulmalik et al., 2017
Bacterial blight, Blast	Xa21 and xa13, Pi54	STS	xa13 prom, pTA 248 and Pi54 MAS	Rice	Arunakumari et al., 2016
Quality protein	opaque2	SSR	phi057 and umc1066	Maize	Kostadinovic et al., 2016
Grain protein con- tent, Thousand grain weight	GPC-B1 and TGW	SSR	Xucw108, Xgwm297	Wheat	Vishwakarma et al., 2016 and 2014
Fusarium head blight	Fhb7, Fhb1	SSR	XsdauK66 and Xcfa2240 (Fhb7), Xgwm493 and Xgwm533 (Fhb1)	Wheat	Guo et al. 2015
Leaf curl disease	Ту-2, Ту-3, Ту-5	Linked markers	Ty-2, Ty-3, Ty-5, qTy10.1	Tomato	Prasanna <i>et al.,</i> 2015
Blast and bacterial blight	Pi9(t), Xa23, tms5	SCAR/ EST/Indel marker	Pb8, C189, IDtms5	Rice	Ni et al., 2015
Rice tungro disease	RTSV	SSR	RM336	Rice	Shim et al., 2015



Table 2:

Conclusion

Marker assisted selection is a technology that has already proved its value. Due to the number of QTLs, genes and markers identified the MAS is likely to become more valuable. Many organizations and private sectors succeed in implementing MAS and produced new lines with desirable traits. But still reduced cost and optimized strategies for integrating MAS with phenotypic selection are needed before the technology can reach its full potential.

References

Abdulmalik R., Menkir A., Meseka S.K., Unachukwu N., Ado S.G., Olarewaju J.D., Aba D.A., Hearne S., Crossa J., Gedil M. (2017). Genetic gains in grain yield of a maize population improved through marker assisted recurrent selection under stress and non-stress conditions in West Africa. Front. Plant Sci., 8: 841.

Acquaah G. (2012). Principles of plant genetics and breeding. 2nd ed. Wiley-Blackwell, Oxford.

Agrios G.N., (2004). Plant pathology. 5th ed. London: Elsevier, 922 p.

Alkimim E.R., Caixeta E.T., Sousa T.V., Pereira A.A., de Oliveira A.C.B., Zambolim L., Sakiyama N.S. (2017). Mol Breeding, 37: 6.

Arunakumari K., Durgarani C.V., Satturu V., Sarikonda K.R., Chittoor P.D.R., Vutukuri B., Laha G.S., Nelli A.P.K., Gattu S., Jamal M., Prasadbabu A., Hajira S., Sundaram R.M. (2016). Marker-Assisted Pyramiding of Genes Conferring Resistance Against Bacterial Blight and Blast Diseases into Indian Rice Variety MTU1010. Rice Sci., 23: 306–316.

Baliyan N., Malik R., Rani R., Mehta K., Vashisth U., Dhillon S., Boora K.S. (2018). Integrating marker-assisted background analysis with foreground selection for pyramiding bacterial blight resistance genes into Basmati rice. C. R. Biol., 341: 1-8.

Barnard A.D., Labuschagne M.T., Van Niekerk H.A. (2002). Heritability estimates of bread wheat quality traits in The Western Cape Province of South Africa. Euphytica, 127: 115–122.

Baum M., Weeden N.F., Muehlbauer F.J., Kahl G., Udupa S.M., Eujay I., Weigand F., Harrabi M., Bouznad Z. (2000). Marker technology for plant breeding. In: Knight R. (eds) Linking Research and Marketing Opportunities for Pulses in the 21st Century. Curr. Plant Sci. Biot. vol 34. Springer, Dordrecht.

Beckmann J.S., Soller M. (1986). Restriction fragment length polymorphisms and genetic improvement of agricultural species, Euphytica, 35: 111-124.

Bernatzky R., Tanksley S.D. (1986). Towards a saturated linkage map in tomatoes based on isozyme and random cDNA sequences. Genetics, 112: 887–898.

Bohn M., Utz H.F., Melchinger A.E. (1999). Genetic similarities among winter wheat cultivars determined on the basis of RFLPs, AFLPs, and SSRs and their use for predicting progeny variance. Crop Sci., 39:228–237.

Botstein D., White R.L., Skolnick M., Davis R.W. (1980). Construction of a genetic linkage map in man using restriction fragment length polymorphisms. Am. J. Hum. Genet., 32: 314-331.

Bouhadida M., Benjannet R., Madrid E., Amri M., Kharrat M. (2013). Efficiency of marker-assisted selection in detection of Ascochyta blight resistance in Tunisian chickpea breeding lines. Phytopathol. Mediterr., 52: 202-211.

Boyer J.S. (1982). Plant productivity and environment. Science, 218:443-448.

Brachi B., Faure N., Horton M., Flahauw E., Vazquez A., Nordborg M. (2010). Linkage and association mapping of Arabidopsis thaliana flowering time in nature. PLoS Genet., 6:e1000940.

Burkhamer R.L., Lanning S.P., Martens R.J., Martin J.M., Talbert L.E. (1998). Predicting progeny variance from parental divergence in hard red spring wheat. Crop Sci., 38:243–248.

Bustamam M., Tabien R.E., Suwarno A., Abalos M.C., Kadir T.S., Ona I., Bernardo M., Veracruz C.M., Leung H. (2002). Asian Rice Biotechnology Network: Improving Popular Cultivars Through Marker-Assisted Backcrossing by the NARES. Poster presented at the International Rice Congress. September 16-20, Beijing, China.

Cao Y., Li S., Wang Z., Chang F., Kong J., Gai J., Zhao T., (2017). Identification of Major Quantitative Trait Loci for Seed Oil Content in Soybeans by Combining Linkage and Genome-Wide Association Mapping. Front Plant Sci., 8: 1222.

Chen L., Li Y.X., Li C., Wu X., Qin W., Li X., Wang T. (2016). Fine-mapping of qGW4.05, a major QTL for kernel weight and size in maize. BMC Plant Biol., 16: 81.

Chung O.K., Ohm J.B., Lookhart G.L. (2003). Quality characteristics of hard winter and spring wheats grown under an Over–Wintering Condition. J. of Cereal Sci., 37:91–99.

Courtois G., Smahi A., Reichenbac J., Döffinger R., Cancrini C., Bonnet M., Casanova J.L. (2003). A hypermorphic IkappaBalpha mutation is associated with autosomal dominant anhidrotic ectodermal dysplasia and T cell immunodeficiency. J. Clin. Invest., 112: 1108–1115.

Cramer G.R. (2010). Abiotic stress and plant responses from the whole vine to the genes. Aust. J. Grape Wine Res., 16: 86-93.

Cramer G.R., Urano K., Delrot S., Pezzotti M., Shinozaki K. (2011). Effects of abiotic stress on plants: a systems biology perspective. BMC Plant Biol., 11:163.

DePauw R.M., Townley-Smith T.F., Humphreys G., Knox R.E., Clarke F.R., Clarke J.M. (2005). Lillian hard red spring wheat. Can. J. Plant Sci., 85: 397-401.

Dimitrijević A., Ivana I., Dragana M., Sandra C., Siniša J., Tijana Z., Zvonimir S. (2017). Oleic acid variation and marker-assisted detection of Pervenets mutation in high- and low-oleic sunflower cross. Crop Breed. Appl. Biot., 17: 235-241.

Dita M.A., Rispail N., Prats E., Rubiales D., Singh K.B. (2006). Biotechnology approaches to overcome biotic and abiotic stress constraints in legumes. Euphytica, 147:1–24.

Dura S., Mahmud A.M., Duwayri A., Miloudi M.N. (2012). Detection of molecular markers associated with yield and yield components in durum wheat (Triticum turgidum L. var. durum Desf.) under drought conditions. African Journal of Agricultural Research, 8:2118-2128.

Gorthy S., Narasu L., Gaddameedi A., Sharma H.C., Kotla A., Deshpande S.P., Are A.K. (2017). Introgression of Shoot Fly (Atherigona soccata L. Moench) Resistance QTLs into Elite Post-rainy Season Sorghum Varieties Using Marker Assisted Backcrossing (MABC). Front Plant Sci., 8: 1494. Gumber R.K., Schill B., Link W., von Kittlitz E., Melchinger A.E. (1999). Mean, genetic variance, and usefulness of selfing progenies from intra- and inter-pool crosses in faba beans (Vicia faba L.) and their prediction from parental parameters. Theor. Appl. Genet., 98:569–580.

Guo J., Zhang X., Hou Y., Cai J., Shen X., Zhou T., Xu H., Ohm H.W., Wang H., Li A., Han F., Wang H., Kong L. (2015). High-density mapping of the major FHB resistance gene Fhb7 derived from Thinopyrum ponticum and its pyramiding with Fhb1 by marker-assisted selection. Theor. Appl. Genet., 128: 2301-16.

Gupta S., Srinagesh D., Rai S.S., Gaur V.K., Priestley K., Du Z. (2001). The thick early Archean crust-Teleseismic results from Dharwar Craton, Eos Trans. AGU, 82(47), Fall Meet. Suppl., S11D-03.

Gururani M.A., Upadhyaya C.P., Strasser R.J., Woong Y.J., Park S.W. (2012. Physiological and biochemical responses of transgenic potato plants with altered expression of PSII manganese stabilizing protein Plant Physiol. Biochem., 58: 182-194.

Halewood M., Chiurugwi T., Hamilton R.S., Kurtz B., Marden E., Welch E., Michiels F., Mozafari J., Sabran M., Patron N., Kersey P., Bastow R., Dorius S., Dias S., McCouch S., Powell W. (2018). Plant genetic resources for food and agriculture: opportunities and challenges emerging from the science and information technology revolution. New Phyto., 217: 1407-1419.

Harzing A.W. (2007). Publish or Perish, available from https://harzing.com/resources/publish-or-perish

Helms T., Orf J., Vallad G., McClean P. (1997). Genetic variance, coefficient of parentage, and genetic distance of six soybean populations. Theor. Appl. Genet., 94:20–26.

Hossain F., Muthusamy V., Pandey N., Vishwakarma A. K., Baveja A., Zunjare R. U., Gupta, H. S. (2018). Marker-assisted introgression of opaque2 allele for rapid conversion of elite hybrids into quality protein maize. Journal of genetics, 97: 287-298.

Hu H., Dai M., Yao J., Xiao B., Li X., Zhang Q., Xiong L. (2006). Overexpressing a NAM, ATAF, and CUC (NAC) transcription factor enhances drought resistance and salt tolerance in rice. Proc. Natl. Acad. Sci.,103: 12987-12992.

Humphreys D.G., Townley-Smith T.F., Lukow O., McCallum B., Gaudet D., Gilbert J., Fetch T., Menzies J., Brown D., Czarnecki E. (2010). Burnside extra strong hard red spring wheat. Can. J. Plant Sci., 90: 79-84.

Hung H.Y., Browne C., Guill K., Coles N., Eller M., Garcia A., Lepak N., Melia-Hancock S., Oropeza-Rosas M., Salvo S., Upadyayula N., Buckler E.S., Flint-Garcia S., McMullen M.D., Rocheford T.R., Holland J.B. (2012). The relationship between parental genetic or phenotypic divergence and progeny variation in the maize nested association mapping population. Heredity, 108: 490–499.

Jaccoud D., Peng K., Feinstein D., Kilian A. (2001). Diversity arrays: a solid state technology for sequence information independent genotyping. Nucleic Acids Research, 29, e25.

Jighly A., Oyiga B.C., Makdis F., Nazari K., Youssef O., Tadesse W., Abdalla O., Ogbonnaya F.C. (2015). Genome-wide DArT and SNP scan for QTL associated with resistance to stripe rust (Puccinia striiformis f. sp. tritici) in elite ICARDA wheat (Triticum aestivum L.) germplasm. Theor. Appl. Genet., 128:1–19.

Jongeneel C.V. (2000). Searching the expressed sequence tag (EST) databases: panning for genes. Briefings in Bioinformatics, 1:76-92.

Karikari B., Li S., Bhat J.A., Cao Y., Kong J., Yang J., Gai J., Zhao T. (2019). Genome-Wide Detection of Major and Epistatic Effect QTLs for Seed Protein and Oil Content in Soybean Under Multiple Environments Using High-Density Bin Map. Int. J. Mol. Sci., 20: 979.

Khan G.H., Shikari A.B., Vaishnavi R., Najeeb S., Padder B.A., Bhat Z.A., Parray G.A., Bhat M.A., Kumar R., Singh N.K. (2018). Marker-assisted introgression of three dominant blast resistance genes into an aromatic rice cultivar Mushk Budji. Sci. Rep., 8:4091.

Körber N., Bus A., Li J., Parkin I.A., Wittkop B., Snowdon R.J., Stich B. (2016). Agronomic and Seed Quality Traits Dissected by Genome-Wide Association Mapping in Brassica napus. Front. Plant Sci., 7, 386.

Kostadinovic M., Ignjatovic-Micic D., Vancetovic J., Ristic D., Bozinovic S., Stankovic G., Mladenovic Drinic S. (2016). Development of High Tryptophan Maize Near Isogenic Lines Adapted to Temperate Regions through Marker Assisted Selection - Impediments and Benefits. PLoS One 11: e0167635.

Krishnakumar R., Kumaravadivel N. (2018). Marker-assisted selection for biotic stress (Bacterial leaf blight and gall midge) tolerance in Bc4F4 generation of rice (Oryza sativa L.). Plant Breeding, 9: 275 – 282.

Kumar A., Sandhu N., Dixit S., Yadav S., Swamy B., Shamsudin N. (2018). Marker-assisted selection strategy to pyramid two or more QTLs for quantitative trait-grain yield under drought. Rice (New York, N.Y.). 11: 35.

Kumar G., Anil R.R., Hanchinal Shrinivas D., Suma B. (2017). Marker Assisted Gene Pyramiding of Leaf Rust Resistance Genes Lr24 and Lr28 in the Background of Wheat Variety DWR 162 (Triticum aestivum L.). Int. J. Curr. Microbiol. App. Sci., 6: 1883-1893.

LeDeaux J.R., Graham G.I., Stuber C.W. (2006). Stability of QTLs involved in heterosis in maize when mapped under several stress conditions. Maydica, 51: 151–167.

Li C.X., Xu W.G., Guo R., Zhang J.Z., Qi X.L., Hu L., Zhao M.Z. (2018). Molecular marker assisted breeding and genome composition analysis of Zhengmai 7698, an elite winter wheat cultivar. Scientific reports, 8: 322.

Li L., Tacke E., Hofferbert H-R., Lubeck J., Strahwald J., Draffehn A.M., Walkemeier B., Gebhardt C. (2013). Validation of candidate gene markers for marker-assisted selection of potato cultivars with improved tuber quality. Theor. Appl. Genet., 126:1039–1052.

Liang F., Deng Q., Wang Y., Xiong Y., Jin D., Li J., Wang B. (2004). Molecular marker-assisted selection for yield-enhancing genes in the progeny of "9311×O. rufipogon" using SSR. Euphytica, 139: 159–165.

Lombardi M., Materne M., Cogan N.O.I., Rodda M., Daetwyler H.D., Slater A.T., Forster J.W., Kaur S. (2014). Assessment of genetic variation within a global collection of lentil (Lens culinaris Medik.) cultivars and landraces using SNP markers. BMC Genet., 15:150.

Lucas P.M. (1992). Health and Disease in the Late Prehistoric Southeast in Verano, J.W. & Ubelaker, D.H. (eds) Disease and Demography in the Americas, Smithsonian Institute Press, Washington.

Ma J., Zhang H., Li S., Zou Y., Li T., Liu J., Ding P., Mu Y., Tang H., Deng M., Liu Y., Jiang Q., Chen G., Kang H., Li W., Pu Z., Wei Y., Zheng Y., Lan X. (2019). Identification of quantitative trait loci for kernel traits in a wheat cultivar Chuannong16. BMC Genet., 20:77.



MacMillan K., Emrich K., Piepho H.P., Mullins C.E., Price AH. (2006). Assessing the importance of genotype \times environment interaction for root traits in rice using a mapping population. I: A soil-filled box screen. Theor. Appl. Genet., 113: 977–986.

Maeda Y., Yamanaka Y., Sasaki A., Suzuk M., Ohtaishi N. (1990). Haematology in sika deer (Cervus nippon yesoensis Heude, 1884). The Japanese Journal of Veterinary Science, 52: 35-41.

Mannur D.M., Babbar. A, Thudi M., Sabbavarapu M.M., Roorkiwal M., Yeri S.B., Bansal V.P., Jayalakshmi S.K., Yadav S.S., Rathore A., Chamarthi S.K., Mallikarjuna P.B., Gaur P.M., Varshney R.K. (2018). Super Annigeri 1 and improved JG 74: two Fusarium wilt-resistant introgression lines developed using marker-assisted backcrossing approach in chickpea (Cicer arietinum L.). Mol. Breeding, 39: 2.

Martin J.M., Talbert L.E., Lanning S.P., Blake N.K. (1995). Hybrid performance in wheat as related to parental diversity. Crop Sci., 35: 104-108.

Melchinger A.E. (1993). Use of RFLP markers for analyses of genetic relationships among breeding materials and prediction of hybrid performance. p. 621–628. In D.R. Buxton et al. (ed.) International crop science I. CSSA, Madison, WI

Melchinger A.E., Utz H.F., Schön C.C. (1998). Quantitative trait locus (QTL) mapping using different testers and independent population samples in maize reveals low power of QTL detection and large bias in estimates of QTL effects. Genetics, 149: 383–403.

Miklas P.N., Kelly J.D., Beebe S.E., Blair M.W. (2006). Common bean breeding for resistance against biotic and abiotic stresses: from classical to MAS breeding. Euphytica, 147:105-131.

Nachit M.M. (1998). Association of grain yield in dryland and carbon isotope discrimination with molecular markers in durum (Triticum turgidum L. var. durum). In: Proceedings of 9th International wheat symposium, Saskatoon, Saskatchewan, Canada, pp. 218-223.

Nachit M.M., Elouafi I. (2004). Durum adaptation in the Mediterranean dryland: breeding, stress physiology and molecular markers. Challenges and Strategies of Dryland Agriculture, 32: 203-218. Crop Science Society of America and American Society of Agronomy, Special Publication 32: 203-218.

Nakashima K., Nakashima Ito Y., Yamaguchi-Shinozaki Y. (2009). Transcriptional regulatory networks in response to abiotic stresses in Arabidopsis and grasses Plant Physiology, 149: 88-95.

Ni D., Songa F., Nia J., Zhangb A., Wang C., Zhaoc K., Yanga Y., Weia P., Yanga J., Li L. (2015). Marker-assisted selection of two-line hybrid rice for diseaseresistance to rice blast and bacterial blight. Field Crops Research, 184: 1–8.

Olson R.K., Wise B., Conners F., Rack J., Fulker D. (1989). Specific deficits in component reading and language skills: Genetic and environmental influences. Journal of Learning Disabilities, 22: 339-348.

Omoigui L.O., Kamara A.Y., Moukoumbi Y.D., Ogunkanmi L.A., Timko M.P. (2017). Breeding cowpea for resistance to Striga gesnerioides in the Nigerian dry savannas using marker-assisted selection. Plant Breeding, 136: 393-399.

Paran I., Michelmore R.W. (1993). Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. Theor. Appl. Genet., 85: 985-993.

Powell WW, Koput K, Smith-Doerr L. (1996). Interorganization collaboration and the locus of



innovation: Networks of learning in biotechnology. Admin. Sci. Quart., 41: 116-145.

Prasanna H.C., Kashyap S.P., Krishna R., Sinha D.P., Reddy S., Malathi V.G. (2015). Marker assisted selection of Ty-2 and Ty-3 carrying tomato lines and their implications in breeding tomato leaf curl disease resistant hybrids. Euphytica, 64: 256–264.

Pukalenthy B., Manickam D., Chandran S., Adhimoolam K., Sampathrajan V., Rajasekaran R., Arunachalam K., Ganapathyswamy H., Chocklingam V., Muthusamy V., Hossain F., Natesan S. (2019). Incorporation of opaque-2 into 'UMI 1200', an elite maize inbred line, through marker-assisted backcross breeding. Biotechnology and Biotechnological Equipment, 33: 144-153.

Randhawa H.S., Asif M., Pozniak C.J., Clarke J.M., Graf R.J., Fox S.L., Humphreys D.G., Knox R.E., DePauw R.M., Singh A.K., Cuthbert R.D., Hucl P.J., Spaner D.M. (2013). Application of molecular markers to wheat breeding in Canada. Plant Breeding, 132: 458-471.

Ribaut J.M., Ragot M. (2007). Marker-assisted selection to improve drought adaptation in maize: the backcross approach, perspectives, limitations, and alternatives. J. Exp. Bot., 58: 351–360.

Ryan J., Ibrahim H., Dakermanji A., Niane A.A. (2012). Training and Capacity Building: An Essential Strategy for Development at an International Research Center. Sustainable Agriculture Research, 1.

Sax K. (1923). The association of size differences with seed-coat pattern and pigmentation in Phaseolus Vulgaris. Genetics, 8:552–560.

Sayed H., Baum M. (2018). Marker-assisted selection for scald (Rhynchosporium commune L.) resistance gene(s) in barley breeding for dry areas. Journal of Plant Protection Research, 4: 335–344.

Shim J., Torollo G., Angeles-Shim R.B., Cabunagan R.C., Choi I.R., Yeo U.S., Ha W.G. (2015). Rice tungro spherical virus resistance into photoperiod-insensitive japonica rice by marker-assisted selection. Breed. Sci., 65:345-51.

Singh I., Smita S., Mishra D.C., Kumar S., Singh B.K., Rai A. (2017). Abiotic Stress Responsive miRNA-Target Network and Related Markers (SNP, SSR) in Brassica juncea. Front. Plant Sci., 8: 1943.

Singh M., Mallick N., Chand S., Kumari P., Sharma J.B., Sivasamy M., Jayaprakash P., Prabhu K.V., Jha S.K. (2017). Marker-assisted pyramiding of Thinopyrum-derived leaf rust resistance genes Lr19 and Lr24 in bread wheat variety HD2733. J. Genet., 96:951–957.

Sourour A., Othmani A., Bechrif S., Rezgui M., Ben Younes M. (2018). Correlation between agronomical and quality traits in durum wheat (Triticum durum Desf.) germplasm in semi arid environment. Advances in Plants and Agriculture Research, 8:612–615

Steele J.G., Sanders A.E., Slade G.D., Allen P.F., Lahti S., Nuttall N., Spencer A.J. (2004). How do age and tooth loss affect oral health impacts and quality of life? A study comparing two national samples. Community Dent. Oral Epidemiol., 32:107-14.

Steele K.A., Price A.H., Shashidhar H.E., Witcombe J.R. (2006). Marker-assisted selection to introgress rice QTLs controlling root traits into an Indian upland rice variety. Theor. Appl. Genet., 112: 208–222.

Tadesse W., Abdalla O., Ogbonnaya F., Nazari K., Tahir I., Baum M. (2012). Agronomic performance of elite stem rust resistant spring wheat genotypes and association among trial sites in



the CWANA region. Crop Sci., 52: 1105-1114.

Tadesse W., Ogbonnaya F.C., Jighly A., Nazari K., Rajaram S., Baum M. (2014). Association Mapping of Resistance to Yellow Rust in Winter Wheat Cultivars and Elite Genotypes. Crop Sci., 54: 607–616.

Tadesse W., Nachit M., Abdalla O., Rajaram S. (2016). "Wheat breeding at ICARDA: Achievements and prospects in the CWANA region", in Alain, B., Bill, A. and Maarten van, G. (Eds), The World Wheat Book Volume 3: A History of Wheat Breeding, Lavoisier, Paris, I

Tanaka Y., Yoneda H., Hosokawa M., Miwa T., Yazawa S. (2014). Application of marker-assisted selection in breeding of a new fresh pepper cultivar (Capsicum annuum) containing capsinoids, low-pungent capsaicinoid analogs. Sci. Hortic., 165: 242–245.

Tanksley S.D., Young N.D., Paterson A.H., Bonierbale M.W. (1989). RFLP mapping in plant breeding: new tools for an old science. BioTechnology, 7: 257-264.

Thoday J.M. (1961). Location of polygenes. Nature, 191: 368-370.

Todaka D., Nakashima K., Maruyama K., Kidokoro S., Osakabe Y., Ito Y., Matsukura S., Fujita Y., Yoshiwara K., Ohme-Takagi M., Kojima M., Sakakibara H., Shinozaki K., Yamaguchi-Shinozaki K. (2012). Rice phytochrome-interacting factor-like protein OsPIL1 functions as a key regulator of internode elongation and induces a morphological response to drought stress. Proc. Natl. Acad. Sci. U.S.A, 109:15947–15952.

Vishwakarma M.K., Mishra V.K., Gupta P.K., Yadav P.S., Kumar H., Joshi A.K. (2016). Introgression of the high grain protein gene Gpc-B1 in an elite wheat variety of Indo-Gangetic Plains through marker assisted backcross breeding. Curr. Plant Biol., 1: 60–67.

Visioni A., Gyawali S., Selvakumar R., Gangwar O.P., Shekhawat P.S., Bhardwaj S.C., Al-Abdallat A.M., Kehel Z., Verma R.P.S. (2018). Genome Wide Association Mapping of Seedling and Adult Plant Resistance to Barley Stripe Rust (Puccinia striiformis f. sp. hordei) in India. Front. Plant Sci., 9: 520.

Vos P., Hogers R., Bleeker M., Reijans M., van de Lee T., Hornes M., Freijters A., Pot J., Peleman J., Kuiper M., Zabeau M. (1995). AFLP: a new concept for DNA fingerprinting. Nucleic Acids Res., 21: 4407-4414.

Wang D, Zhao L, Li K, Ma Y, Wang L, Yang Y, Yang Y, Zhi H (2017). Marker-assisted pyramiding of soybean resistance genes RSC4, RSC8, and RSC14Q to soybean mosaic virus. J. Integr. Agr., 16: 2413–2420.

Welcker C., Boussuge B., Benciveni C., Ribaut J.M., Tardieu F. (2007). Are source and sink strengths genetically linked in maize plants subjected to water deficit? A QTL study of the responses of leaf growth and anthesis-silking interval to water deficit. J. Exp. Bot., 58:339–349.

Wilde F., Schön C.C., Korzun V., Ebmeyer E., Schmolke M., Hartl L., Miedaner T. (2008). Theor. Appl. Genet., 117: 29.

Williams J.G.K., Kubelik A.R., Livak K.J., Rafalski J.A., Tingey S.V. (1990). DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. Nucl. Acids Res., 18: 6531-6535.

Xu Y., Crouch J.H. (2008). Marker-assisted selection in plant breeding: from publication to practice. Crop Sci., 48: 391-407.



Xu Y., Zhang X.Q., Harasymow S., Westcott S., Zhang W., Li C. (2018). Molecular marker-assisted backcrossing breeding: an example to transfer a thermostable β -amylase gene from wild barley. Mol. Breeding, 38:63.

Yagdi K.S. (2009). Heritability, variance components and correlations of yield and quality traits in durum wheat (Triticum durum desf.). Pak. J. Bot., 41:753–759.

Yamaguchi-Shinozaki K., Shinozaki K. (2006). Transcriptional regulatory networks in cellular responses and tolerance to dehydration and cold stresses. Annu. Rev. Plant Biol., 57: 781–803.

Yang D., Tang J., Yang D., Chen Y., Ali J., Mou T. (2019). Improving rice blast resistance of Feng39S through molecular marker-assisted backcrossing. Rice (New York, N.Y.) 12: 70.

Young N.D. (1999). A cautiously optimistic vision for marker-assisted breeding. Mol. Breeding, 5: 505-510.

Yugander A., Sundaram R.M., Singh K., Ladhalakshmi D., Subba Rao L.V., Madhav M.S., Badri J., Prasad M.S., Laha G.S. (2018. Incorporation of the novel bacterial blight resistance gene Xa38 into the genetic background of elite rice variety Improved Samba Mahsuri. PLoS One,13:e0198260.

Zhao Q.Y., Zhu Z., Zhang Y.D., Zhao L., Chen T., Zhang Q.F., Wang C.L. (2008). Correlation analysis between genetic distance of SSR markers and heterosis japonica. The Fifth National Congress of Plant Molecular Breeding-cum-academic exchanges Proceedings.

Zhao X., Tan G., Xing Y., Wei L., Chao Q., Zuo W., Lübberstedt T., Xu M. (2012). Marker-assisted introgression of qHSR1 to improve maize resistance to head smut. Mol. Breeding, 30:1077–1088.

References